

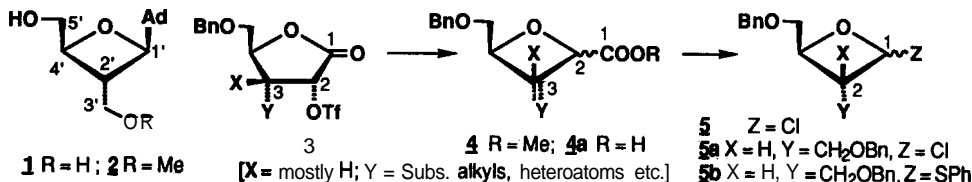
RING CONTRACTION REACTIONS OF **2-O-METHANESULFONATES** OF α -**HYDROXY- γ -LACTONES** IN AQUEOUS MEDIUM TO **OXETANE-2-CARBOXYLIC** ACIDS: A CONVENIENT SYNTHESIS OF **3'-O-METHYLOXETANOCIN** AND A FORMAL SYNTHESIS OF **OXETANOCIN**

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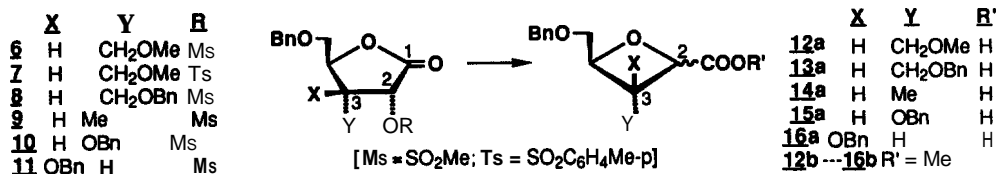
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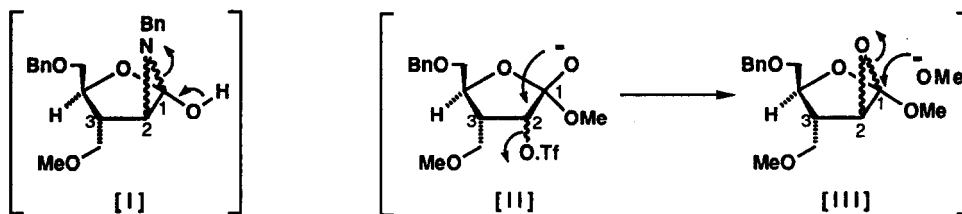
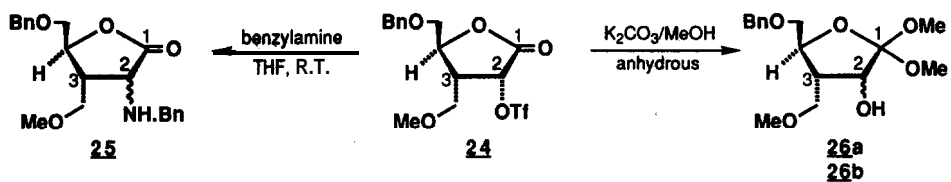
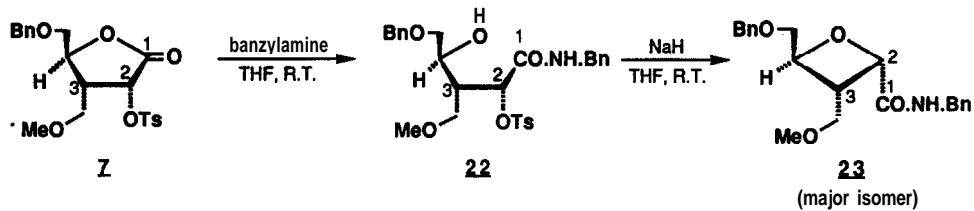
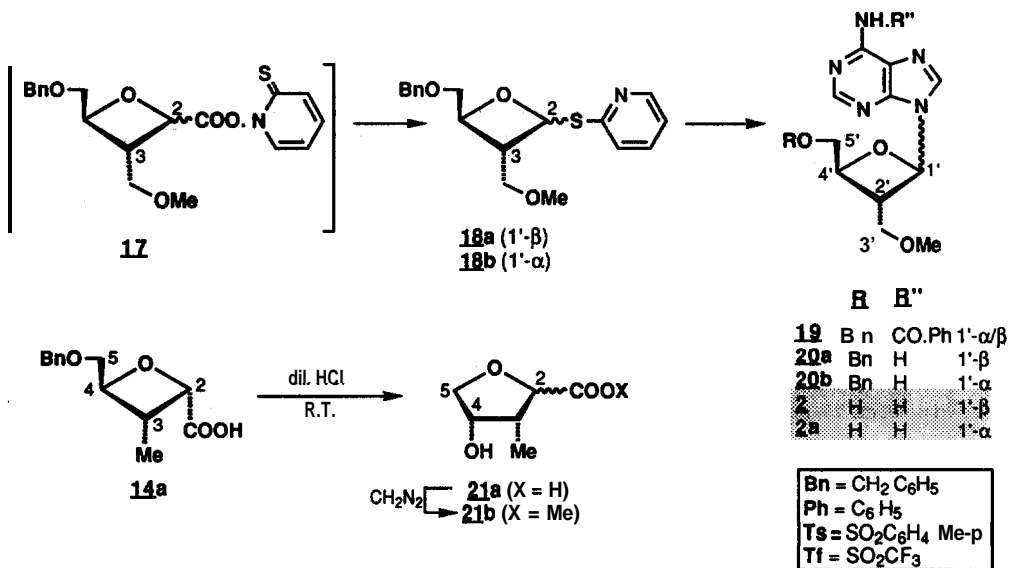
Abstract: Barton decarboxylative rearrangement¹⁵ of thiohydroxamic ester 17 directly provided the key **oxetanosyl-thiopyridyl** glycosides **18** which were successfully coupled to N-benzoyladenine. A two-step ring contraction of the tosylate 7 to the oxetane-2-carboxamide 23 is described. Attempted ring contraction of the triflate 24 gave the unexpected products 25 and 26. A ring expansion reaction (e.g. **14a** \rightarrow 21a) was also observed.

Oxetanocin **1**, the first natural oxetanosyl-N-glycoside and its analogs show promising antiviral activities against HSV, humancytomegalovirus and HIV.^{1,2} A number of elegant syntheses of **1** and other analogs have been reported.³⁻⁶ In a versatile approach, the **2-O-triflate- γ -lactones** **3** were ring contracted to oxetane-2-carboxylic esters **4** in the presence of anhydrous K_2CO_3 /methanol.⁶ After hydrolysis the resulting acids **4a** were then subjected to Barton decarboxylative chlorination¹⁵ to provide unstable chlorooxetanes **5**.^{6b,7} Thus, in a "low yield" synthesis of **1** (Ad = adenine), the chlorooxetane **5a** was converted to protected **1** and its α -anomer.^{6b} For the success of the transformations **3** \rightarrow **4**, anhydrous conditions were found to be essential. It was also shown that under these conditions, the corresponding **2-O-mesyates** provided virtually no oxetanes.^{6a,b} We now show that especially under aqueous hydrolytic conditions, ring contraction of such **2-O-mesyates** is an efficient process culminating in an opportune synthesis of **2** and its anomer **2a**.



The **lactone** substrates **6** - **11** and **24** were readily prepared from diacetone glucose according to published procedures.^{6b,8} In initial experiments, when the mesylate **6** and the tosylate **7** were treated with anhydrous K_2CO_3 in methanol over 24-36 hours, the oxetane-2-methylesters **12b** were isolated in ~30% and 10% yields respectively (anomer **ratio** ~1:1); the unchanged lactones being the major components in these reactions.^{9,10} This suggested that the poor leaving group ability of **O-mesyate** and **tosylate** imposed an element of reversibility causing the intermediate oxy-anion to relactonize. In contrast, we found that treatment of the mesylates **6** and **8** - **11** in methanol with aqueous sodium hydroxide readily provided¹¹ the **oxetane-2-carboxylic acids** **12a** (80%), **13a** (72%), **14a** (56%) **15a** (48%) and **16a** (35%). The predominant isomers in these reactions were derived from inversion of configuration at C-2 (**5:1** for **12a** - **14a** and **3:2** for **15a** and **16a**).^{13,14} These oxetane-2-carboxylic acids were fully characterized as their methyl esters **12b** - **16b**.





The **carboxylic acid 12a** was converted in a one-pot sequence to the **thiohydroxamic** ester 17 which was subjected to photolytic decarboxylative rearrangement^{15,16} to provide thiopyridyl glycosides **18a/b** in 30% yield.¹⁷ Coupling of **18a/b** with **N-benzoyladenine** (4 equiv.) in DMF, in the presence of bromine (4 equiv.) and 4A molecular sieves,¹⁸ led to protected nucleoside (anomeric) mixture 19 in ~60% yield. Removal of the **N-benzoyl** group (**NaOMe, MeOH**) provided easily separable¹⁹ anomers **20a**, m.p. 107-110° (**H₁-α, d, J_{1,2}**: 5.9 Hz) and **20b**, m.p. 134-136° (**H₁-β, d, J_{1,2}**: 6.96 Hz) in over 90% yield (**20a:20b, 2:3**). Both isomers were debenzylated (Pd black, **EtOH, ~quant.** yield) to provide 3'-O-methyl oxetanocin 2, m.p. 203-205°, [α]_D-14.3° (**CHCl₃**), and its l'-epimer 2a.²⁰ This also constitutes a formal synthesis of oxetanocin 1 since the **oxetane-2-** methylcarboxylate 13b²¹ has been previously converted to **1.6b**. We found that **oxetane-2-carboxylic** acids 12a – 16a on either prolonged storage or in contact with added acids undergo a ring expansion reaction (e.g. **14a → 21a**; -30% yield).²² Such furanosyl-C-glycosides could also be detected immediately after some of the ring contraction reactions described above.²³ Ring expansion reactions of N-benzoyl oxetanocin diesters have been observed during transglycosidation attempts.²⁴ Formation of 218 appears to be an unusual example of a ring expansion reaction accompanying debenzylation. We have not studied these ring contraction reactions in the presence of other protecting groups.

In a two-step variant of the above ring contraction reactions, the tosylate 7 upon treatment with benzylamine in THF at room temperature cleanly gave the acyclic benzylamide 22. Treatment of 22 with **NaH** in THF at room temperature (-2.5 hrs.) led to **oxetane-2-carboxamide** 23 in overall 90% yield from 7 (anomer ratio **5:1**; δ 5.04, d, **J_{2,3}** 6.6 Hz, **H₂-β**, major isomer; δ 4.64, d, **J_{2,3}** 6.0 Hz, **H₂-α**, minor isomer).

Reaction of the more reactive triflate 24 with benzylamine as above gave exclusively the α -aminobenzyl- γ -lactone 25 rather than 23 – the ring contraction product.²⁵ Also, attempted ring contraction of 24 with **K₂CO₃** in anhydrous methanol²⁶ did not provide 12b but only the orthoesters **26a/b** (-60% yield).²⁷ In view of ample precedents for ring contraction of closely related triflates,^{6b} these anomalous results are inexplicable at this time.

Perhaps an 'aziridine-aminal' species such as [I] might be involved in the formation of 25 although direct nucleophilic displacement of the triflate group by benzylamine is a more plausible explanation. We propose an 'epoxy-acetal' species [III] as a possible intermediate in the formation of **26a/b**. The tetrahedral intermediate [I I] may act as a common species for the well documented ring contraction reactions," as well as **26a/b** via [I I I]. To the best of our knowledge, formation of **26a/b** from 24 is a novel orthoester forming reaction under basic conditions.

We are currently evaluating the scope and limitations of the above ring contraction reactions as well as the unusual transformations, **14a → 218** and **24 → 26a/b**.²⁸

Acknowledgements: We thank Sir Derek Barton. FRS, for stimulating discussions. We would also like to thank Dr. B. Pramanik and Mr. P. Banner for high resolution mass spectral data.

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7. These chlorooxetanes 5 were so unstable that it was found necessary to first convert them to thiophenyl glycosides **5b** for purification; they were then regenerated back to chlorooxetanes for coupling with N-benzoyladenine just before use.^{6b} The thiopyridylglycosides **18a/b** used here were stable to chromatography and to storage at -10° under argon for several months.

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9. All new compounds were characterized by high resolution mass spectra, ^1H and ^{13}C NMR spectra. When necessary 2D (^1H - ^1H) and 2D (^1H - ^{13}C) COSY NMR spectra were obtained. Elemental analyses were obtained for crystalline compounds only. Yields refer to isolated products and have not been optimized. Selective spectral data is given here.
10. 6: ^1H NMR [400 MHz, CDCl_3] δ 2.95 (m, 1H), 3.25 (s, 3H), 3.35 (s, 3H), 3.55 (m, 3H), 3.75 (dd, J 10.8, 2.2 Hz, 1H), 4.55 (ABq, J 12.1 Hz, 2H), 4.7 (s, broad, 1H), 5.68 (d, J_{2,3} 9 Hz, 1H), 7.2-7.4 (m, 5H).
12b: ^1H NMR [CDCl_3] δ 4.9 (d, J_{2,3} 6.0 Hz, 2 α -H); 5.14 (d, J_{2,3} 8.5 Hz, 2 β -H). C₂-isomer ratio, ~1:1.
11. Typical procedure: A 10% methanolic solution of the mesylate was treated with aq. 1 N NaOH (3.5 equiv.) at R.T. After ~15 hrs. the reaction mixture was cooled (ii bath) and neutralized with 1 N HCl (3.5 equiv.). Evaporation of the solvents *in-vacuo* at 35-40°C followed by extraction of the residue with CH_2Cl_2 provided the crude product which was chromatographed on silica gel; eluent: 12 • 25% acetone in n-hexane followed by 25% methanol in CH_2Cl_2 . Both isomers could be separated in this manner although mixtures were used for Barton decarboxylative rearrangement.¹⁵
12. 12a: ^1H NMR [CDCl_3] δ 3.3 (m, 1H), 3.38 (s, 3H), 3.5 (dd, J 11.5, 1.5 Hz, 1H), 3.58 (m, 2H), 3.8 (dd, J 11.5, 2.6 Hz, 1H), 4.7 (ABq, J 12 Hz, 2H), 4.87 (m, 1H), 4.9 (d, J_{2,3} 6.6 Hz, 2 α -H, minor isomer), 7.3-7.4 (m, 5H); δ 3.3 (s, 3H), 3.4 (m, 1H), 3.5 (m, 2H), 3.7 (m, 2H), 4.65 (ABq, J 12 Hz, 2H), 4.93 (m, 1H), 5.1 (d, J_{2,3} 9 Hz, 2 β -H, major isomer), 7.3-7.4 (m, 5H).
13. For ^1H coupling constants in oxetanes, see: A. Balsamo, C. Ceccarelli, P. Crotti and F. Macchia, *J. Org. Chem.*, **1975**, **40**, 473.
14. In the formation of minor retention products, the available evidence neither rules out interconversion of lactone sulfonate esters nor epimerization at some latter stage. More detailed studies would be necessary to fully understand these results.
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16. Thiiohydroxamic ester 17 was prepared *in-situ* via mixed anhydride procedure by addition of isobutyrylchloroformate and N-methyl morpholine to a 10% solution of 12a in dry THF at -20°C. After 5 minutes activation period a solution of N-hydroxy pyridinethione and triethylamine in dry THF was added. After stirring for ~30 minutes, the reaction was allowed to warm to room temperature and irradiated with 150 watts tungsten lamp for ~60 minutes. Work-up by extractive isolation with ethylacetate provided the crude product, which was chromatographed on silica gel (eluent: 25% ethylacetate in n-hexane) to provide pure 18a and 18b in 30% combined yield. Some of the less polar fractions also contained the isobutyryl ester of the acid 12a (~25%).
17. 18a: ^1H NMR [CDCl_3] δ 63.2 (m, 1H), 3.4 (s, 3H), 3.6 (dd, 2H), 3.75 (d, broad, 2H), 4.65 (m, 3H), 4.85 (m, 1H), 6.86 (d, J_{1,2} 6.6 Hz, H₁- α , minor isomer), 7.0-7.66 (m, 8H), 8.45 (m, 1H); 18b: δ 3.38 (s, 3H), 3.6-3.8 (m, 4H), 4.65 (m, 3H), 4.85 (m, 1H), 6.88 (d, J_{1,2} 7.4 Hz, H₁- β , major isomer), 7.0-7.6 (m, 8H), 8.45 (m, 1H).
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19. Silica gel chromatography (TLC grade), eluent: 1% methanol in ethylacetate.
20. 2: ^1H NMR [CD_3OD] δ 63.4 (s, 3H), 3.66 (dd, J 5.2, 2H), 3.76 (dd, J 12.3 Hz, 1H), 3.9 (m, 2H), 4.68 (m, 1H), 6.5 (d, J_{1,2} 6.07, H₁- α , minor anomer), 8.2 (s, 1H) and 8.66 (s, 1H); 2a: δ 2.95 (s, 3H), 3.31-3.43 (m, 4H), 3.61 (m, 1H), 3.77 (dd, J 12.5, 3.8 Hz, 1H), 3.89 (dd, J 12.5, 2.9 Hz, 1H), 4.98 (m, 1H), 6.7 (d, J_{1,2} 6.88 Hz, H₁- β , major anomer), 8.2 (s, 1H), 8.46 (s, 1H).
21. 13b: ^1H NMR [CDCl_3] δ 53.5 (m, 1H), 3.6 (dd, 2H), 3.67 (s, 3H), 3.69 (dd, 2H), 4.47 (dd, 3H), 4.65 (dd, 2H), 4.88 (m, 1H), 6.14 (d, J_{2,3} 8.8 Hz, H₂- β , major isomer), 7.2-7.4 (m, 10H); minor isomer: δ 4.92 (d, J_{2,3} 6.8 Hz, H₂- α).
22. 14b: ^1H NMR [CDCl_3] δ 1.1, (d, 3H), 3.4 (m, 1H), 3.64 (dd, 2H), 3.8 (s, 3H), 4.45-4.7 (m, 3H), 5.11 (d, J_{2,3} 6.9 Hz, H₂- β), 7.2-7.4 (m, 5H); signals at δ 1.3 (d, 3H), 2.94 (m, 1H), 3.7 (s, 3H) and 4.65 (d, J_{2,3} 6.0 Hz, H₂- α) indicating the presence of ~20% retention product (by integration).
21b: ^1H NMR [CDCl_3] δ 1.22 (d, 1.22 (d, J 7 Hz, 3H), 2.22 (m, broad, 1H), 2.29 (m, 1H), 3.76 (s, 3H), 3.92 (dd, J 10, 1 Hz, 1H), 4.12 (dd, J 10, 1H), 4.15 (d, J 10 Hz, 1H) and 4.28 (dd, 4, 4 Hz, 1H).
23. These observations appear to be related to some recent examples of tetrahydrofuran cyclizations with concomitant debenzylations, see: H. Dahmlow, J. Mulzer, C. Seitz, A. Ft. Strecker and A. Kohlmann, *Tetrahedron Lett.*, **1992**, **33**, 3607.
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25. 25: ^{13}C NMR [CDCl_3] δ 176.8 (s), 139.8 (s), 137.8 (s), 126.5 (d), 128.3 (d), 127.9 (d), 127.8 (d), 127.2 (d), 127.1 (d), 76.8 (d, C₄), 73.7 (t), 70.6 (t), 70.1 (t), 53.1 (l) and 44.3 (d, C₃); ^1H NMR [CDCl_3] δ 2.47 (m, 1H), 3.26 (s, 3H), 3.41 (dd, 12.5 Hz, 1H), 3.43 (dd, J 11, 2 Hz, 1H), 3.50 (d, J 10, H₂), 3.6 (dd, J 11, 4 Hz, 1H), 3.77 (dd, J 11, 2 Hz, 1H), 3.94 (d, J 12, 1H), 4.0 (d, J 12, 1H), 4.55 (d, J 11 Hz, 1H), 4.58 (d, J 11 Hz, 1H). This data also ruled out the ring contracted oxetane-amide structure 23.
26. To a 10% solution of the triflate 24 in dry methanol was added K_2CO_3 (anhydrous; 6 equiv.) in one portion. The reaction mixture was stirred at room temperature for 30 minutes and filtered to remove solids. Evaporation of the filtrate *in-vacuo* and preparative TLC on silica gel (eluent: 40% ethylacetate in n-hexane) provided 26a and 26b as colorless gums.
27. 24: ^1H NMR [CDCl_3] δ 62.95 (m, 1H), 3.35 (s, 3H), 3.54 (m, 3H), 3.75 (d, 1H), 4.55 (dd, 2H), 4.67 (s, broad, 1H), 5.78 (d, J_{2,3} 8.8 Hz, 1H), 7.2-7.4 (m, 5H).
26a: ^{13}C NMR [CDCl_3] δ 138.2 (s), 128.3 (d), 127.6 (d), 127.5 (d), 120.8 (s, C₁), 79.1 (d, C₄), 73.2 (t), 72.5 (d, C₂), 69.7 (t), 59.0 (q), 50.9 (q), 49.3 (q), and 44.0 (d, C₃); 26b: δ 137.8 (s), 128.4 (d), 127.7 (d), 119.3 (s, C₁), 77.8 (d, C₄), 73.9 (d, C₂), 73.5 (t), 72.4 (t), 71.8 (t), 58.9 (q), 50.0 (q), 49.5 (q) and 48.4 (d, C₃); Stereochemistry at C₂ in 26a: δ H₃ 2.45, δ C₃ 44.0 (OH function is *cis* to H₃) end in 26b: δ H₂ 2.30, δ C₃ 48.4 (OH function is *trans* to H₃)
IR [neat, isomers 26a and 26b]: no carbonyl absorption.
28. A. K. S. would like to dedicate this contribution to his mentor, Dr. R. H. B. Gall.